

## SHORT REPORTS

### GUAIANOLIDES FROM *GOCHNATIA SMITHII*\*

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**Key Word Index**—*Gochnattia smithii*, Compositae, sesquiterpene lactones, guaianolides, 8 $\alpha$ -acetoxydehydrocostus lactone, 8 $\alpha$ -tigloyloxydehydrocostus lactone

**Abstract**—The structure and stereochemistry of 8 $\alpha$ -acetoxy and 8 $\alpha$ -tigloyloxy dehydrocostus lactones, two guaianolides isolated from the aerial parts of *Gochnattia smithii*, were established by chemical and spectroscopic methods

#### INTRODUCTION

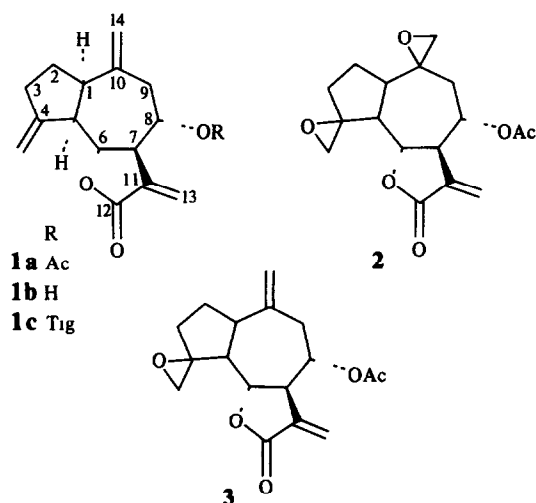
From the large genus *Gochnattia* (tribe Mutisieae, Compositae) only five species have been analyzed chemically [1-4]. Although all of them contain triterpenes, sesquiterpene lactones have been found only in *G. discoidea* [2], *G. rusbyana* [3] and *G. paniculata* [4]. From the last two zaluzanin C and dimeric guaianolides were also isolated. We now report the isolation of two guaianolides from *Gochnattia smithii* Robins & Greem, both of them closely related to dehydrocostus lactone derivatives isolated from Vernoniaceae tribe [5, 6].

#### DISCUSSION

The major component, compound A, C<sub>17</sub>H<sub>20</sub>O<sub>4</sub> (MS), mp 120-121°, [ $\alpha$ ]<sub>D</sub> + 87.3° (c 0.179, CHCl<sub>3</sub>), is a sesquiterpene lactone of the guaianolide type and it was formulated as **1a**. This compound showed absorption bands in the IR spectrum at 1768 cm<sup>-1</sup> ( $\gamma$ -lactone- $\alpha,\beta$ -unsaturated), 1735 cm<sup>-1</sup> (saturated ester), 1653 and 1640 cm<sup>-1</sup> (double bonds). The <sup>1</sup>H NMR spectrum revealed the presence of three terminal methylenes, one of them corresponding to that conjugated with the  $\gamma$ -lactone ( $\delta$  6.18, *d*, *J* = 3.5 Hz and 5.61, *d*, *J* = 3.5 Hz). The other two methylenes were located at C-14 (5.01, *s br* and 4.89, *s br*) and C-15 (5.24, *s br* and 5.06, *s br*). The signal for the proton under the lactone oxygen bridge appeared as a doublet of doublets at 3.98 (*J* = 10, 9 Hz), its multiplicity and coupling constants in agreement with a *trans*- $\gamma$ -lactone closed to C-6. At higher field one singlet for an acetate methyl group was evident (2.11, *s*, 3H). Although the signal for the proton under this group was overlapped with the signals of the terminal methylene protons, this functionality was tentatively located at C-8. In order to sweep out the methylene signals compound A (**1a**) was epoxidized. In the <sup>1</sup>H NMR spectrum of the diepoxy derivative (**2**) the H-8 signal was clearly observed at  $\delta$  5.14 (*J* = 10, 5.5 and 4.5 Hz). The

values of the coupling constants indicated the  $\beta$ -orientation for H-8, therefore the C-8 acetoxy group must be  $\alpha$ . From this reaction also the monoepoxy-derivative (**3**) was obtained. Its <sup>1</sup>H NMR spectrum showed that only the signals for the C-15 protons were shifted to high field ( $\delta$  3.31 and 2.88, AB).

The acetyl moiety was eliminated by saponification of **1a** producing **1b** in poor yield. In the IR spectrum the carbonyl band for the ester function did not appear, instead, a hydroxyl absorption (3430 cm<sup>-1</sup>) was observed. The <sup>1</sup>H NMR spectrum showed the H-8 signal shifted to higher field ( $\delta$  3.89) and partially superimposed with the H-6 signal. The physical constants for this compound (**1b**) are almost identical with those of 8 $\alpha$ -hydroxydehydrocostus lactone obtained from *Centaurea canariensis* [7, 8] [they only differ in the small additional coupling of the C-13 protons ( $\delta$  6.25, *dd*, *J* = 3.5, 1 Hz and 6.14, *dd*, *J* = 3, 1 Hz)]. The above mentioned facts permitted us to establish the structure of compound A as 8 $\alpha$ -acetoxydehydrocostus lactone (**1a**).



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Table 1  $^1\text{H}$  NMR spectral data of compounds 1–3 (80 MHz,  $\text{CDCl}_3$ , TMS as internal standard, values in parentheses are coupling constants in Hz)

|       | 1a                        | 1b                         | 1c                                                                     | 2                                 | 3                        |
|-------|---------------------------|----------------------------|------------------------------------------------------------------------|-----------------------------------|--------------------------|
| H-6   | 3.98 <i>dd</i><br>(10, 9) | 3.95 <i>dd</i><br>(10, 9)  | 4.03 <i>dd</i><br>(10, 9)                                              | 4.20 <i>dd</i><br>(11, 9)         | 4.1 <i>dd</i><br>(11, 9) |
| H-7   | 3.11 <i>m</i>             | *                          | 3.18 <i>m</i>                                                          | 3.12 <i>m</i>                     | *                        |
| H-8   | 4.8–5.1*                  | 3.90*                      | 4.9–5.2*                                                               | 5.14 <i>ddd</i><br>(10, 5.5, 4.5) | 4.99*                    |
| H-13  | 6.18 <i>d</i><br>(3.5)    | 6.25 <i>dd</i><br>(3.5, 1) | 6.18 <i>d</i><br>(3.5)                                                 | 6.24 <i>d</i><br>(3.5)            | 6.2 <i>d</i><br>(3.5)    |
| H-13' | 5.61 <i>d</i><br>(3)      | 6.14 <i>dd</i><br>(3, 1)   | 5.59 <i>d</i><br>(3)                                                   | 5.61 <i>d</i><br>(3)              | 5.58 <i>d</i><br>(3)     |
| H-14  | 5.01 <i>sbr</i>           | 5.04 <i>br</i>             | 5.04 <i>br</i>                                                         | 2.79 AB (2H)                      | 5.13 <i>br</i>           |
| H-14' | 4.89 <i>sbr</i>           | 4.95 <i>br</i>             | 4.92 <i>br</i>                                                         |                                   | 4.93 <i>br</i>           |
| H-15  | 5.24 <i>sbr</i>           | 5.27 <i>br</i>             | 5.28 <i>br</i>                                                         | 3.31 <i>d</i><br>(5)              | 3.31 <i>d</i><br>(5)     |
| H-15' | 5.06 <i>sbr</i>           | 5.06 <i>br</i>             | 5.1 <i>br</i>                                                          | 2.90 <i>d</i><br>(5)              | 2.88 <i>d</i><br>(5)     |
| OCOR' | 2.11 <i>s</i> (3H)        |                            | 6.92 <i>qbr</i><br>(7.5)<br>1.92 <i>dbr</i><br>(7.5)<br>1.79 <i>br</i> | 2.12 (3H)                         | 2.14 (3H)                |

\*Superimposed signal

The minor component, compound B,  $\text{C}_{20}\text{H}_{24}\text{O}_4$  (EM), mp 96–98°,  $[\alpha]_D + 109.2^\circ$  (c 0.152,  $\text{CHCl}_3$ ) obtained from the less polar fraction of the chromatography, also is an  $\alpha, \beta$ -unsaturated- $\gamma$ -lactone (1768  $\text{cm}^{-1}$ ). The  $^1\text{H}$  NMR spectrum indicates a great similarity with that of compound A. Compound B is esterified with a tigloyl residue instead of the acetyl of 1a. This was evident from the presence in the  $^1\text{H}$  NMR spectrum of a broad quartet at  $\delta$  6.92 and two broad signals (3H each) at 1.92 and 1.79. The IR absorption band at 1713  $\text{cm}^{-1}$  coupled with the mass peaks at  $m/z$  228  $[\text{M} - 100]^+$ , 83 (100) and 55 (11.5) confirmed this assumption and allowed us to assign the structure of 8 $\alpha$ -tigloyloxydehydrocostus lactone (1c) for compound B, which is identical with ferreyanthus lactone [9].

## EXPERIMENTAL

Aerial parts (3.3 kg) of *Gochnatia smithii* (voucher on deposit in the herbarium of the Instituto de Biología, U N A M, MEXU 342416) were extracted with  $\text{CHCl}_3$ . The crude gum (84 g) was percolated through a Tonsil column [10]. 11 fractions were collected. Fractions 1–8 ( $\text{CHCl}_3$ ), 9–14 (EtOAc). Fractions 7–12 (18.4 g) were chromatographed over 550 g of silica gel (Merck 60, 70–230 mesh), 250 ml fractions were collected. Fractions 27–49 ( $\text{CHCl}_3$ ) gave, after crystallization from EtOAc–hexane, 79.6 mg of compound B (1c) mp 96–98°,  $[\alpha]_D + 109.2^\circ$  (c 0.152,  $\text{CHCl}_3$ ), IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  1768, 1713, 1653 (calc for  $\text{C}_{20}\text{H}_{24}\text{O}_4$ , MW 328). Found MW (EIMS) 328. Other significant peaks in the MS were at  $m/z$  228  $[\text{M} - \text{C}_5\text{H}_4\text{O}_2]^+$ , 213  $[\text{M} - \text{Me}]^+$ , 83 (100), 55 (11.5).

Fractions 50–94 ( $\text{CHCl}_3$ ) gave 3.6 g of compound A (1a) mp 120–121° (EtOAc–hexane),  $[\alpha]_D + 87.3^\circ$  (c 0.179,  $\text{CHCl}_3$ ), IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  1768, 1735, 1652, 1640 (calc for  $\text{C}_{17}\text{H}_{20}\text{O}_4$ , MW 288). Found MW (EIMS) 288. Other relevant peaks in the

MS were at  $m/z$  246  $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ , 213  $[\text{M} - \text{HOAc}]^+$ , 213  $[\text{M} - \text{Me}]^+$ , 91 (33.8), 43 (100).

**Epoxidation of 1a** MCPBA (82 mg) was added to a soln of 1a (68.4 mg) in  $\text{CHCl}_3$  (8 ml). The mixture was oxidized at room temp for 40 hr. The soln was diluted with  $\text{CHCl}_3$ , washed with satd  $\text{NaHCO}_3$  soln and  $\text{H}_2\text{O}$ , dried and evaporated. The residual gum exhibited two spots on TLC. It was separated by prep TLC on silica gel (Me<sub>2</sub>CO–hexane, 3:7,  $\times 3$ ). The substance with  $R_f$  0.4 (28 mg) was 3 mp 148–151° (Me<sub>2</sub>CO–hexane), IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  1762, 1735, 1645, 1638 (calc for  $\text{C}_{17}\text{H}_{20}\text{O}_5$ , MW 304). Found MW (EIMS) 304. Other significant peaks in the MS were at  $m/z$  262  $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ , 244  $[\text{M} - \text{HOAc}]^+$ , 229  $[\text{M} - \text{Me}]^+$ , 91 (100), 43 (69.7). The substance 2 with  $R_f$  0.25 (21 mg) crystallized from Me<sub>2</sub>CO–hexane, mp 238–240°, IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  1765, 1738, 1655 (calc for  $\text{C}_{17}\text{H}_{20}\text{O}_6$ , MW 320). Found MW (EIMS) 320. Other peaks in the MS were at  $m/z$  290  $[\text{M} - \text{CH}_2\text{O}]^+$ , 278  $[\text{M} - \text{C}_2\text{H}_2\text{O}]^+$ , 260  $[\text{M} - \text{HOAc}]^+$ , 240  $[\text{M} - \text{CH}_2\text{O} - \text{C}_2\text{H}_2\text{O}]^+$ , 230  $[\text{M} - \text{CH}_2\text{O}]^+$ , 91 (73), 43 (100).

**Hydrolysis of 1a** Compound 1a (92.9 mg) was added to a soln of KOH (184 mg) in  $\text{H}_2\text{O}$  (10 ml). The mixture was left to stand at 60° for 1 hr with stirring. The soln was neutralized with 10%  $\text{H}_2\text{SO}_4$  soln, saturated with NaCl and extracted with EtOAc. The washed and dried extract was evaporated and the solid residue was crystallized from Me<sub>2</sub>CO–*i*-propyl ether–hexane to provide 30.4 mg of 1b, mp 106–107°,  $[\alpha]_D + 64.7^\circ$  (c 0.295,  $\text{CHCl}_3$ ), IR  $\nu_{\text{max}}^{\text{CHCl}_3} \text{ cm}^{-1}$  3430, 1765, 1658, 1645 (calc for  $\text{C}_{15}\text{H}_{18}\text{O}_3$ , MW 246). Found MW (EIMS) 246. Other MS peaks were at  $m/z$  228  $[\text{M} - \text{H}_2\text{O}]^+$ , 213  $[\text{M} - \text{Me}]^+$ , 91 (100).

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## HELIANGOLIDES AND BEJARANOLIDES FROM *CONOCLINIOPSIS PRASIIFOLIA*

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**Key Word Index**—*Conocliniopsis prasifolia*, Compositae, sesquiterpene lactones, furoheliangolides, bejaranolides

**Abstract**—A reinvestigation of the aerial parts of *Conocliniopsis prasifolia* afforded two furoheliangolides, conoprasiolide-9-*O*,5'-*O*-diacetate and 5'-desoxyconoprasiolide, as well as two bejaranolides, 9 $\beta$ -hydroxy-4*E*-bejaranolide and 3 $\alpha$ ,9 $\beta$ -dihydroxy-4*E*-bejaranolide. The structures were elucidated by  $^1\text{H}$  NMR spectroscopy. The chemistry of *Conocliniopsis* supports the proposed close relationship to *Bejaranoa*.

### INTRODUCTION

The investigation of the new monotypic Brazilian genus *Conocliniopsis* (tribe Eupatorieae, subtribe Gyptidinae) [1] afforded furoheliangolides similar to those present in the closely related genus *Bejaranoa* [2, 3]. The reinvestigation of the polar fractions of the aerial parts of *Conocliniopsis prasifolia* (DC) K et R gave two further furoheliangolides and two bejaranolides which strongly supported the close relationship of this genus to *Bejaranoa* [4]. The results will be discussed in this paper.

### RESULTS AND DISCUSSION

The aerial parts of *Conocliniopsis prasifolia* afforded in addition to conoprasiolide (1) [2] and its 5'-*O*-acetate 2 [2] two further derivatives, the tiglate 3 and conoprasiolide-9-*O*,5'-*O*-diacetate (4). The structure of 3 followed from the molecular formula and the  $^1\text{H}$  NMR spectral data (Table 1) which were close to those of 1 [2] and 2 [2]. The difference in the nature of the ester groups in 1 and 3 at C-8 caused some small variations of chemical shifts, especially of H-6 and H-8, while the nature of the ester residue at C-8 clearly followed from the typical signals of a tiglate. The diacetate 4 already was prepared

by acetylation of 1 and 2 [2]. Accordingly, the observed  $^1\text{H}$  NMR spectral data (Table 1) were identical with those reported previously [2].

Furthermore, minute amounts of two additional sesquiterpene lactones were isolated. The less polar lactone showed no molecular ion in the mass spectrum. But as the  $^1\text{H}$  NMR spectrum (Table 1) clearly displayed the characteristic signals of a tiglate, the observed peak at  $m/z$  278 (corresponding to  $\text{C}_{15}\text{H}_{18}\text{O}_5$ ) most likely was formed by elimination of tiglic acid. Spin decoupling of the  $^1\text{H}$  NMR spectrum allowed the assignment of all signals which showed a marked similarity to the spectrum of 4*E*-bejaranolide [3]. However, the H-9 signals were replaced by a double doublet at  $\delta$  3.6. Irradiation of this signal collapsed the doublet at 2.29 to a singlet and the signal at 5.86 to a doublet. The latter was further coupled with the fourfold doublet at 2.85. Its irradiation caused the expected changes of the H-13 signals as well as that of H-6, and therefore was due to H-7. Addition of deuterium oxide collapsed the double doublet at 4.36 to a doublet while the doublet at 2.29 disappeared, thus indicating a hydrogen bonded hydroxy group at C-9, similar to the situation in 1 and 3. Accordingly, the stereochemistry at C-9 also was the same and all data agreed with the proposed structure 5.